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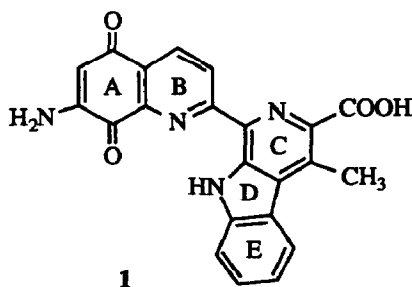
A Straightforward and Practical Formal Synthesis of Lavendamycin Ethyl Ester.

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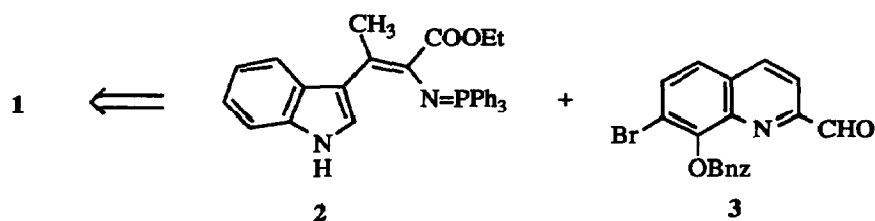
Abstract. *Aza Wittig reaction of iminophosphorane derived from ethyl α -azido- β -(3-indolyl)butyrate and 8-benzyloxy-7-bromo-2-formylquinoline is the key step in a new synthesis of lavendamycin ethyl ester.*

Lavendamycin **1**, isolated in 1981 from the fermentation broths of *Streptomyces lavendulae*,¹ displays an important antimicrobial activity, inhibitory activity for the lysogenic strain of *E. coli* and more importantly antitumor² activity against P-388 and L-1210 and is an inhibitor of the HIV reverse transcriptase.³ Consequently, this target molecule has been the focus of synthetic efforts since its initial structural identification. Synthetic approaches towards **1** involve either Bichler-Napijeralski reaction between β -methyltryptophan and quinaldic acid derivatives⁴ or Pictet-Spengler cyclization reaction between β -methyltryptophan and 8-benzyloxy-2-formylquinoline⁵ to build the ring C. The Boger approach⁶ is based on the formation of the ring B by Friedlander cyclization between an *ortho*-aminobenzaldehyde possessing suitable functionalities and a 1-acetyl- β -carboline derivative, available by regioselective inverse electron demand [4+2] cycloaddition reaction of electron deficient 1,2,4-triazine with α -arylenamine, followed by palladium (0)-mediated β -carboline synthesis. A recent formal synthesis of lavendamycin methyl ester involves a modified Knoevenagel-Stobbe pyridine formation and further ring D construction by thermolytic nitrene insertion.⁷



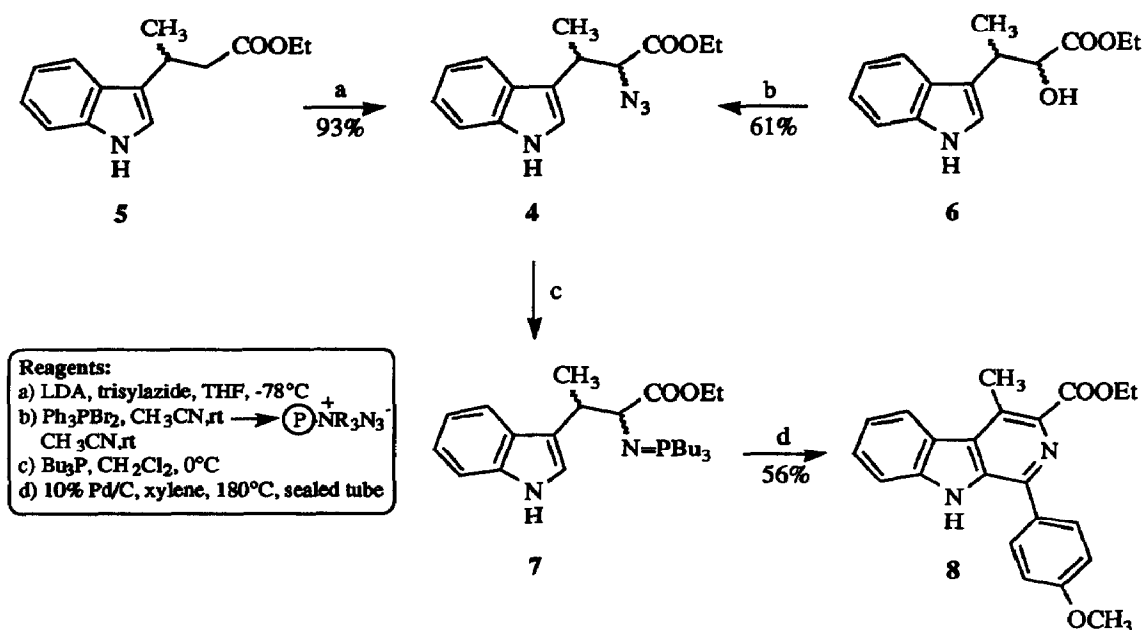
The tandem aza Wittig/electrocyclic ring-closure has been successfully utilized for the synthesis of several types of carboline derivatives.⁸ Model studies starting from the iminophosphorane derived from the ethyl α -azido- β -(3-indolyl)propenoate resulted in the formation of 1-(2-quinolyl)- β -carbolines with contain the gross ring system of lavendamycin.⁹ We now report a slight modification of this strategy in the form of an efficient and practical formal synthesis of lavendamycin ethyl ester. At first the iminophosphorane **2** was selected in order to incorporate in the β -carboline ring the functionalities necessary for preparation of the natural product into the cyclization step. Unfortunately, attempts to prepare **2**: condensation of 3-acetylindole with ethyl

azidoacetate in the presence of different bases and dehydration agents; Michael-addition of indole on ethyl α -azidobutenoate and Heck reaction of 3-iodoindole with ethyl α -azidobutenoate in the presence of palladium reagents, were unsuccessful. Keeping this mind, we turned our attention to the preparation of the dihydroderivative ethyl α -azido- β -(3-indolyl)butyrate **4**. This compound was prepared by two ways¹⁰ a) starting from the readily available ethyl 3-(3-indolyl)butyrate¹¹ **5** by the imide-enolate azidation procedure described by Evans and co-



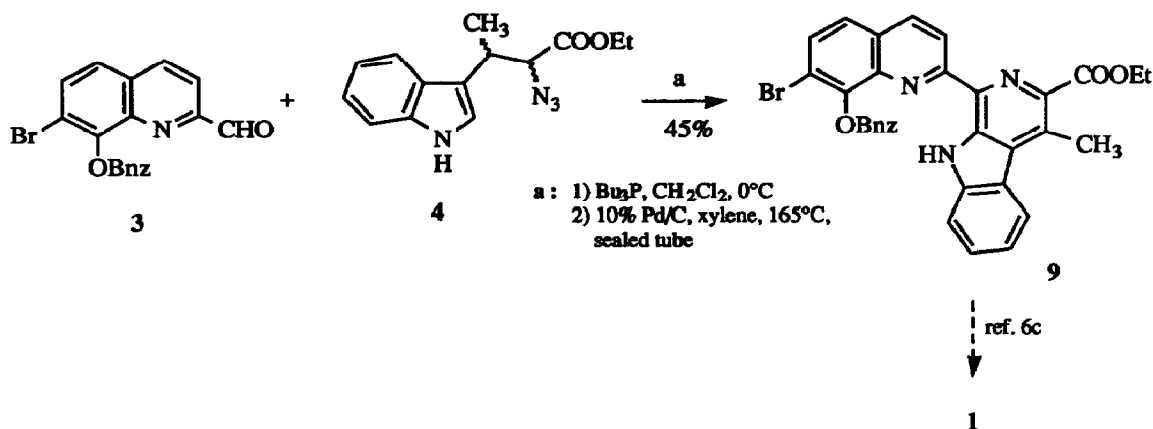
workers¹² using LDA/2,4,6-triisopropylbenzenesulfonyl azide¹³ (trisylozide) or b) starting from 2-hydroxy-3-(3-indolyl)butyrate **6**, available from indole and ethyl 2,3-epoxybutyrate,¹⁴ by sequential treatment with triphenylphosphine dibromide and polymeric quaternary ammonium azide.¹⁵

The azide **4** does not convert to the corresponding iminophosphorane by treatment with triphenylphosphine even after an extended reaction time (10h). However, treatment of **4** in dichloromethane at 0°C with tributylphosphine under nitrogen produces the iminophosphorane **7**, which could be isolated as a viscous oil in almost quantitative yield. In a model study this iminophosphorane reacted with aromatic aldehydes, namely 4-methoxy benzaldehyde to give the β -carboline derivative **8**, possessing suitable functionalities in the pyridine ring. Best results were obtained when the reaction was carried out in xylene in a sealed tube at 180°C in the presence of Pd/C.



The next step was the preparation of the aldehyde **3** which possesses suitable functionality for conversion to the 7-aminoquinoline-5,8-quinone, AB ring of lavendermycin; This compound was easily prepared in high yield by a two-step sequence starting from 2-amino-3-benzyloxy-4-bromobenzaldehyde^{6b}: a) Friedlander condensation with acetone provided 8-benzyloxy-7-bromo-3-methylquinoline in 78%, b) oxidation with selenium dioxide afforded **3** (mp 79-81°C) in 82% yield.¹⁶ Reaction of the aldehyde **3** with the iminophosphorane **7**, generated in situ from **4** and tributylphosphine, in a sealed tube at 165°C in the presence of Pd/C provided **9** in 45% yield.¹⁷

This constitutes a formal total synthesis of lavendamycin ethyl ester, since **9** is the ethyl ester of the Boger lavendamycin intermediate,^{6c} and it may be converted into the final product in a straightforward manner.



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- Typical procedure:* To a solution of diisopropylamine (0.082 g, 8.1 mmol) in dry THF (30 ml), was added dropwise n-butyllithium (2M, 3.9 ml, 7.8 mmol). The solution was stirred at -78°C under dry N_2 for 45 min. A solution of ethyl 3-(3-indolyl)butyrate **5** (0.3 g, 1.3 mmol) in THF (15 ml) was added in one portion, and the resultant solution was warmed to -30°C for 15 min and was then recooled to -78°C . Hexamethylphosphoric triamide (HMPA) (3.5 ml, 20 mmol) was added and to the above enolate solution was added a pre-

cooled (-78°C) solution of trisylazide (1.6 g, 5.2 mmol) in dry THF (20 ml). The reaction mixture was stirred at -78°C for 1 h, and then was quenched with glacial acetic acid (1.7 ml, 29.7 mmol). The resulting mixture was allowed to warm to room temperature, stirred for 12h, treated with a saturated solution of NaHCO₃ (100 ml) and extracted with dichloromethane (3x40 ml). The organic extracts were combined, washed with a solution of sodium chloride (1x10 ml), dried (Na₂SO₄) and evaporated in vacuo. The residue was chromatographed on a silica gel column using Et₂O/hexane (1:1) as eluent to afford **4** in 93% yield. ¹H n.m.r. (300 MHz, CDCl₃) δ 1.14 (t, 3H, ³J= 7.1 Hz), 1.43 (d, 3H, ³J= 6.9 Hz), 3.73 (quintuplet, 1H, J= 6.9 Hz, H-β), 4.12 (q, 2H, ³J= 7.1 Hz), 4.21 (d, 1H, J=5.7 Hz, H-α), 7.1 (d, 1H, J= 2.4 Hz, H-2), 7.1-7.3 (m, 2H, H-5 + H-6), 7.36 (dd, 1H, J= 8.1 and 0.8 Hz, H-7), 7.64 (dd, 1H, J= 7.7 and 0.8 Hz, H-4), 8.2 (s, 1H, NH). ¹³C n.m.r. (75 MHz, CDCl₃) δ 13.9 (CH₃), 16.1 (CH₃), 33.4 (C-β), 61.6 (CH₂), 67.2 (C-α), 111.3 (C-7), 116.6 (C-3), 118.7 (C-4), 119.5 (C-5), 122.0 (C-6), 122.2 (C-2), 126.2 (C-3a), 136.2 (C-7a), 169.0 (C=O).

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16. **3**: (82%). ¹H n.m.r. (300 MHz, CDCl₃) δ 5.56 (s, 2H, PhCH₂O), 7.5-7.3 (m, 3H, 2H_m + H_p), 7.51 (d, 1H, J= 8.7 Hz, H-5), 7.67 (dd, 2H, J=7.4, 1.5 Hz, 2H_p), 7.79 (d, 1H, J= 8.7 Hz, H-6), 8.04 (d, 1H, J= 8.4 Hz, H-4), 8.28 (d, 1H, J=8.4 Hz, H-3), 10.23 (s, CHO); ¹³C n.m.r. (75 MHz, CDCl₃) δ 76.8 (PhCH₂O), 117.5 (C-3), 118.0 (C-7), 123.8 (C-6), 128.2 (C_p), 128.3 (2C_p), 128.6 (2C_m), 130.7 (C-4a), 133.5 (C-5), 137.0 (C-8a), 137.8 (C-4), 143.1 (C_p), 151.9 (C-8), 153.0 (C-2), 193.3 (CHO).
17. *Typical Procedure*: To a cooled at 0°C solution of tributylphosphine (0.35 g, 1.7 mmol) in dry dichloromethane (25 ml), was added a solution of **4** (0.47 g, 17 mmol) in the same solvent (20 ml). The mixture was stirred at 0°C under nitrogen for 6 h. The solvent was removed under vacuo and to the residual iminophosphorane **7** were added o-xylene (20 ml), 10% Pd/C (0.051 g), and 8-benzyloxy-7-bromo-2-formyl quinoline **3** (0.6 g, 1.7 mmol). The resultant mixture was treated in a sealed tube at 165°C for 20 h. After cooling, the solution was filtered and the filtrate was concentrated to dryness. The residue was chromatographed on a silica gel column using ethyl acetate/hexane as eluent to give **9** (45%, yellow prisms, m.p. 186-188°C, from dichloromethane/diethyl ether). ¹H n.m.r. (300 MHz, CDCl₃) δ 1.55 (t, 3H, J= 7.2 Hz, CH₃CH₂OCO), 3.10 (s, 3H, CH₃), 4.55 (q, 2H, J= 7.2 Hz, CH₃CH₂OCO), 5.25 (s, 2H, PhCH₂O), 6.27 (d, 1H, J=7.7 Hz, H-8), 7.3-7.2 (m, 2H, H-7 + H-6), 7.4-7.3 (m, 3H, 2H_m + H_p), 7.44 (d, 1H, J= 8.7 Hz, H-5'), 7.62-7.58 (dd, 2H, J= 7.2, 1.8 Hz, 2H_p), 7.62 (d, 1H, J= 8.7 Hz, H-6'), 8.16 (d, 1H, J= 8.7 Hz, H-4'), 8.18 (d, 1H, J= 7.5 Hz, H-5), 8.77 (d, 1H, J= 8.7 Hz, H-3'), 11.65 (s, 1H, NH); ¹³C n.m.r. (75 MHz, CDCl₃) δ 14.6 (qt, ¹J= 126.9, ²J= 2.6 Hz, CH₃CH₂OCO), 16.8 (q, ¹J= 128.5 Hz, CH₃), 61.3 (tq, ¹J= 147.5, ²J= 4.5 Hz, CH₃CH₂OCO), 75.4 (t, ¹J= 146.1, ³J= 4 Hz, PhCH₂O), 112.7 (dd, ¹J= 163.0, ³J= 7.3 Hz, C-8), 117.5 (d, ²J= 12.5 Hz, C-7'), 119.7 (d, ¹J= 169.2 Hz, C-3'), 120.4 (dd, ¹J= 160.6, ³J= 7.1 Hz, C-6), 121.9 (q, ³J= 5.9 Hz, C-4a), 123.4 (dd, ¹J= 159.9, ³J= 7.5 Hz, C-6), 124.6 (dd, ¹J= 164.7, ²J= 4.6 Hz, C-5'), 127.6 (d, ¹J= 159.1 Hz, C_p), 127.8 (d, ¹J= 161.2 Hz, 2C_p), 128.2 (t, ²J= 8.3 Hz, C-4'a), 128.3 (dt, ¹J= 160.1, ²J= 7.5 Hz, C-7), 128.9 (dd, ¹J= 160.1, ²J= 6.6 Hz, 2C_m), 129.9 (d, ²J= 7 Hz, C-4b), 130.7 (d, ¹J= 168.1 Hz, C-6'), 132.0 (q, ²J= 6.2 Hz, C-4), 134.4 (s, C-8'a), 135.4 (s, C-9a), 136.6 (dd, ¹J= 163.2, ²J= 4.5 Hz, C-4'), 137.1 (m, C-8a), 137.3 (m, C_p), 140.6 (q, ³J= 6 Hz, C-3), 142.3 (t, ³J= 6 Hz, C-1), 151.6 (d, ³J= 7 Hz, C-8'), 157.5 (d, ²J= 7.5 Hz, C-2'), 167.6 (s, C=O).

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